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# The Mechanism Of Action Of Polyphenol On Changes In The Dynamics Of Calcium In The Synaptosomes Of The Rat Brain Against The Background Of Glutamate

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#### ABSTRACT

The manuscript shows a short data used using fluorescent probes to study the effect of polyphenol PC-7 on changes in the dynamics of intracellular  $Ca^2$ + content in rat brain synaptosomes, depending on the site of glutamate binding on calcium channels by a specific mediator with glutamate. To measure the amount of cytosolic  $Ca^2$ + synaptosomes, we calculated using the Grinkevich equation. It has been shown that polyphenol PC-7 binds to the  $\beta$ 1-subunit of the voltage-gated calcium channel and allosterically changes its conformation so that the conductivity for  $Ca^2$ + ions increases through the channel, the blocking effect of polyphenol PC-7 can be explained by its binding to voltage-gated calcium channels and activating them.

#### **KEYWORDS**

NMDA-receptors, synaptosomes, intracellular Ca<sup>2+</sup>, glutamate.

## **INTRODUCTION**

Glutamate is the most common mediator in the brain and activates six different classes of receptors, three of which are ionotropic receptors [1]. Each class has its own pharmacological and functional features. There were no significant differences in the pharmacological properties of pre- and postsynaptic glutamate receptors. According to the names of agonists that cause specific physiological responses, N-Methyl-D-aspartate (NMDA) receptors, a-amino-3-hydroxyl-5methyl-4-isoxazolepropionic acid (AMPA) and kainate are isolated. All three classes of glutamate receptors are found in the central nervous system, and in many parts of the brain, the receptors coexist.

Two main types of glutamate receptors (ionotropic and metabotropic) include three functionally distinct receptor classes. Each of the receptors in the class consists of individual subunits encoded by different genes. Subunits of ionotropic glutamate receptors form ion channels that are permeable to cations. Metabotropic glutamate receptors are G-protein-linked receptors, the activation of which regulates the synthesis or degradation of intracellular secondary mediators [1,2,3,4,5,6,7,8].

Receptors differ in the composition of subunits and affinity for glutamate - for NMDA it is  $1 \mu M$ . The functional significance of postsynaptic glutamate receptors is the transfer of excitation between neurons, whereas presynaptic receptors modulate the release of glutamate and other central nervous system mediators. The ligand-activated channel is a tetrameric complex; various combinations of subunits form functionally distinct receptors. The transmembrane topology of the glutamate receptor differs from that of the nicotinic receptor. The polypeptide contains three transmembrane domains  $M_1$ ,  $M_2$  and  $M_4$ [2,9,10,11,12,13]. The NH<sub>2</sub>-end is directed extracellularly; the COOH-end is intracellular. The  $M_2$  domain forms a loop-pin, which forms the internal pore of the channel. The configuration is similar to the K-channels, which implies the common origin of the two classes of ion channels. Ionic channels of glutamate receptors are permeable to Na and Ca ions, but ionic selectivity depends on the subunit composition of the receptor IMPACT FACTOR 2021: 5. 634 OCLC - 1121105553

[2,14,15,16]. Because of the specific nature of nerve cells, a fatal role in them is played by an increase in the concentration of Ca<sup>2+</sup> in the presynaptic region, which leads to a massive release of various neurotransmitters, among which the exciting mediator glutamic acid has a special role. This is due to the fact that, first of all, the most synapses in the brain (about 40%) are glutamatergic and, secondly, the majority of postsynaptic glutamate receptors (NMDA- and AMPA-receptors) control the calcium channels and, accordingly, further increase the concentration Intracellular Ca<sup>2+</sup>. In connection with this positive feedback, the cytoplasmic Ca<sup>2+</sup> concentration of in glutamate-sensitive neurons increases dramatically. Therefore, up to the present time one more name of the processes occurring in the ischemic focus of the brain continues to exist: "exciting toxicity" or otherwise "glutamate excitotoxicity".

The NMDA-receptor is one of the most finely regulated receptors: there are at least six binding sites for endogenous ligands that affect the probability of opening an ion channel. These include activating binding sites with two different agonists, glutamate and glycine, a polyamine binding site, and sites for binding to Mg, Zn, and H. The oxidativereduction status of the receptor also affects conductivity. One of the three pairs of cysteine residues can be both restored (which increases the currents through the channel) and is oxidized with the formation of disulfide bridges (which reduces the currents).

The unique nature of the NMDA-receptor is that its activation requires simultaneous binding of two different agonists, so glutamate and glycine are called "coagents" of the NMDAreceptor. Low molecular weight glycine analogs, including serine and alanine, also act as agonists of the glycine site. In both cases, Disomers are more effective than L-forms. So Dserine, formed by serine racemase, is a strong endogenous agonist of the glycine site. Extracellular Mg ions block the current through the NMDA-receptor channel. The magnesium block is removed when the membrane is depolarized. Protons reduce the frequency of opening the channel, and at pH ~ 6.0, the activation of the receptor is completely suppressed. Binding of polyamines - spermine or spermidine - removes the proton block and, thus, enhances the activation of the receptor. At high concentrations, polyamines, however, also cause a potential-dependent ion channel block [17,18,19,20].

In the central nervous system, Ca<sup>2+</sup> is a ubiquitous cell messenger that participates in the regulation of neurotransmitter release and nerve excitability, and is closely associated with cellular differentiation and migration, synaptic plasticity, neurite growth, and neuronal apoptosis [21,22,23].

Previous studies have shown that in the experiments carried out, the effect of various polyphenols on the change in the dynamics of calcium in the synaptosomes of the rat brain, polyphenols act as antioxidant and neuroprotective properties [24,25,26,27,28,29,30].

Based on this research, in our next scientific experiments, in which synaptosomes of the rat brain were used, loaded with a probe on the cytosolic  $Ca^{2+}$  -Fura-2-AM, it was shown that in physiologically significant concentrations of 10<sup>-7</sup> -10<sup>-5</sup>M on the concentration of cytosolic calcium affects (the polyphenolic compound is usually referred to as (PC-7), isolated from plants Rhus typhina). The mechanism of action of PC-7 on calcium homeostasis has been studied in more detail against the background of glutamate.

#### MATERIAL AND METHODS

Experiments were conducted on 20 outbred male albino rats weighing (200-250 g) contained in a standard vivarium ration. All experiments were performed in accordance with the requirements of "the World Society for the Protection of Animals" and "European Convention for the protection of experimental animals" [31]. Synaptosomes isolated from rat brain by a two-step centrifugation [32]. The whole procedure of selection was carried out at 4°C. To measure the amount of membranebound Ca2+ to synaptosomes placed in a medium similar to that used for isolating the cells, but without apyrase and MgCl<sub>2</sub>, 20  $\mu$ M of chlortetracycline (CTC) was added. Incubated 60 min. To achieve maximum interaction of CT with membrane-bound Ca<sup>2+</sup>, both on the plasma and intracellular membranes. The excitation wavelength of CTC is 405 nm; the registration is 530 nm. The results were expressed as a percentage, taking as 100% the difference between the maximum value of the fluorescence intensity (the fluorescence of the dye saturated with Ca<sup>2+</sup>) and its minimum value (fluorescence of the indicator in the absence of Ca<sup>2+</sup>) obtained after the addition of EGTA.

To measure the amount of cytosolic  $Ca^{2+}$   $[Ca^{2+}]_{in}$  was calculated by the Grinkevich equation [33] in synaptosomes isolated from the brain of rats. To measure the free cytosolic  $Ca^{2+}$  synaptosomes (1 × 108 cells / ml), 4 µM acetoxymethyl ester Fura-2AM was loaded for 40 min at 37 ° C. In this case, the ether group separates from the dye molecules penetrated

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into the cytoplasm by the action of intracellular esterases, resulting in the formed anion Fura-2, which binds Ca2+. After completion of the loading, the dye remaining in the medium was washing removed double by and centrifugation in a standard medium. In the experiments, the concentration of cells in the cell was 5  $\times$  10 <sup>6</sup> cells / ml. Fluorescence excitation was induced at 337 nm, and fluorescence detection at 496 nm. The fluorescence of the dye  $(F_{max})$  saturated with  $Ca^{2+}$  was determined by adding 50  $\mu$ M digitonin to Fura-2AM loaded cells. Fmin was determined by measuring the fluorescence intensity in a non-calcium medium,  $F_{min} = [(F_{max}-F_{af})/3] + F_{af}$ , where  $F_{af}$  is the autofluorescence of cells determined by adding 0.1 mM MnCl<sub>2</sub> to a synaptic loaded with Fura-2AM and treated with digitonin. Registration of changes in the dynamics of calcium in cells used USB-2000 spectrometer (Ocean optics, USA 2010)

### **STATISTICAL ANALYSIS**

The measurements were made using a universal spectrometer (USB-2000). Statistical significance of differences between control and experimental values determined for a number of data using a paired t-test, where the control and the experimental values are taken together, and unpaired t-test, if they are taken separately. The value of P <0.05 indicated a statistically significant differences.

The results obtained are statistically processed to Origin 7,5 (Origin Lab Corporation, USA).

#### **RESULTS AND DISCUSSION**

In our experiments with synaptosomes loaded with a fluorescent probe on Ca<sup>2+</sup>, PC-7 increases

the concentration of intracellular Ca<sup>2+</sup> in a dosedependent manner, both in the calcium-free and calcium-containing media. The action of PC-7 in a noncalcium medium indicates that it stimulates the release of calcium from intracellular compartments. Glutamate reduces the effect of PC-7, which may indicate that part of the external calcium comes under the influence of PC-7 through potentialsensitive calcium channels (Fig.1.A, B).

The effects of glutamate have been studied in rat brain synaptosomes. The effect of glutamate was observed depolarization of the synaptic membrane and an increase in intracellular calcium without a noticeable change in the concentration of internal sodium ions. An increase in the synaptosomal calcium and a decrease in the electrical potential of transmembrane cells was prevented by the addition of glutamate. Activation of the glutamate receptor causes the opening of calcium channels, the influx of calcium into the synaptosomes, and the depolarization of the synaptosomal plasma membrane, followed by the release of amino acid neurotransmitters (Fig.1.C).

Glutamate partially reduces the effect of PC-7, which may indicate that part of the external calcium comes under the influence of PC-7 also through the open glutamine site and in place of the calcium channels of the NMDA-receptors.

Even the preliminary addition of glutamate does not completely eliminate the effects of PC-7, which may indicate that PC-7 has several mechanisms of action on rat brain neurons, the result of which is an increase in intracellular calcium (Fig.1.D).









Figure. 1. The effect of PC-7 and glutamate on the fluorescence of synaptic membranes from the brain of rats pre-loaded with Fura-2. In a calcium-free and calcium-containing medium.

The ordinate shows fluorescence in percent, taking as 100% the difference between the maximum fluorescence intensity value in the presence of 1 mM Ca<sup>2+</sup>, 25  $\mu$ M digitonin and 1 mM rotenone and its minimum value when 1 mM EGTA is added to this medium.

Thus, our experiments in conjunction with the data indicate that PC-7 binds to the  $\alpha_{1}$ - subunit of the potential-controlled calcium channel and allosteric changes its conformation so that the conductivity for Ca<sup>2+</sup> ions increases through the channel.

In the medium with calcium, its effect is somewhat higher, but this excess is eliminated in the presence of glutamate. The effective concentrations of PC-7 are in the range  $10^{-6}$ - $10^{-10}$ 

<sup>5</sup>M. Thus, the blocking effect of PC-7 can be explained by its binding to potential-controlled calcium channels and their activation.

#### CONCLUSIONS

• In this study, calcium chloride changes in synaptosomes were used to study, by means of fluorescent calcium probes, depending on the site of glutamate binding on calcium channels by specific mediators with glutamate. The results indicate a PC-7 binds to the  $\alpha_{1^-}$  subunit of a potential-controlled calcium channel and allosteric changes its conformation so that through the channel the conductivity for Ca<sup>2+</sup> ions increases, blocking effect of PC-7 can be explained by its binding to potential-

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controlled calcium channels and their activation

 As in any field of fundamental science, the completion of one stage does not solve all problems. In what areas can we continue to develop the results and conclusions of this paper. First, it is possible to continue screening among BASs of new substances acting on potential-controlled calcium channels. Secondly, it is necessary to continue the study of substances for which effects on these channels are found: polyphenols and alkaloids.

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## REFERENCES

- Siegel G.J., Albers R.W., Brady, S.T., Price, D.L. Basic Neurochemistry // 7<sup>th</sup> Ed. Academic Press. - 2006. - 992 p.
- Magazannik LG The structure and functions of central and neuromuscular synapses // from the book. The modern course of classical physiology by Ed. Yu.V. Natochina and VA Tkachukayu - M.: Geotar-Media, 2007. P.59-99.
- Olanow C.W., Stern M.B., Sethi K. The scientific and clinical basis for the treatment of Parkinson's disease //

Neurology. — 2009. — V. 72, 21, Suppl. 4. — 149 p.

- Song, A., Charles, A., Koay, S. et al. Volumetric two-photon imaging of neurons using stereoscopy (vTwINS). Nat Methods 14, 420–426 (2017). https://doi.org/10.1038/nmeth.4226
- 5. Siegfried Weisenburger and Alipasha Vaziri. A Guide to Emerging Technologies for Large-Scale and Whole-Brain Optical Imaging of Neuronal Activity. Annual Review of Neuroscience. 2018. 41:1, 431-452
- Velasco M, Rojas-Quintero J, Chávez-Castillo M, Rojas M, Bautista J, et al. (2017) Excitotoxicity: An Organized Crime at The Cellular Level. J Neurol Neurosci. Vol. 8 No. 3: 193
- Auer RN (1986) Progress review: Hypoglycemic brain damage. Stroke 17: 699-708.
- Vincent P, Mulle C (2009) Kainate receptors in epilepsy and excitotoxicity. Neuroscience 158: 309-323.
- 9. Fujikawa DG (2005) Prolonged seizures and cellular injury: understanding the connection. Epilepsy Behav 3: S3-S11.
- 10. Ward RJ, Lallemand F, De Witte P (2009) Biochemical and
- neurotransmitter changes implicated in alcohol-induced brain damage in chronic or 'binge drinking' alcohol abuse. Alcohol Alcohol 44: 128-35.
- Cotter DR, Pariante CM, Everall IP (2001) Glial cell abnormalities in major psychiatric disorders: The evidence and implications. Brain Res Bull 55: 585-595.
- Olney JW (2003) Excitotoxicity, apoptosis and neuropsychiatric disorders. Curr Opin Pharmacol 3: 101-109.

- Leitch B (1992) Ultrastructure of electrical synapses: review. Electron Microsc Rev 5: 311-339.
- Dani A, Huang B, Bergan J, Dulac C, Zhuang X (2010) Super-resolution imaging of chemical synapses in the brain. Neuron 68: 843-856.
- Blakely R, Edwards R (2012) Vesicular and plasma membrane transporters for neurotransmitters. Cold Spring Harb Perspect Biol 4: a005595.
- Ramakrishnan NA, Drescher MJ, Drescher DG (2012) The SNARE complex in neuronal and sensory cells. Mol Cell Neurosci 50: 58-69.
- Kew JN, Kemp JA (2005) Ionotropic and metabotropic glutamate receptor structure and pharmacology. Pyschopharmacology (Berl). 179 : 4-29.
- Palmer LM, Shai AS, Reeve JE, Anderson HL, Paulsen O, et al. (2004) NMDA spikes enhance action potential generation during sensory input. Nat Neurosci 17: 383-90.
- Moriyama Y, Yamamoto A (2004) Glutamatergic chemical transmission: Look! here, there, and anywhere. J Biochem 135: 155-163.
- Berridge MJ (1997) Elementar y and global aspects of calcium signalling. J Physiol 499:291–306
- 22. Bootman MD, Collins TJ, Peppiatt CM et al (2001) Calcium signalling-an overview.Semin Cell Dev Biol 12:3–10
- 23. Bucurenciu I, Bischofberger J, Jonas P (2010) A small number of open Ca2+ channels trigger transmitter release at a central GABAergic syn-apse. Nat Neurosci 13:19–21
- 24. Khushwant S. Bhullar, H. P. Vasantha Rupasinghe, "Polyphenols: Multipotent Therapeutic Agents in Neurodegenerative

Diseases", Oxidative Medicine and Cellular Longevity, vol. 2013, Article ID 891748, 18 pages, 2013.

https://doi.org/10.1155/2013/891748

- 25. Torre, E. Molecular signaling mechanisms behind polyphenol-induced bone anabolism. Phytochem Rev 16, 1183–1226 (2017). https://doi.org/10.1007/s11101-017-9529-x
- 26. Khoshimov N.N., Nasirov K.E. Action of Cytisinum on the Transport Mediators and Calcium Channel of Glutamatergic Neurotransmitter Systems of the NMDA Receptor. European Journal of Medicine.
  2017, 5(2): 56-63. DO-10.13187/ejm.2017.2.31
- 27. Vauzour D. Dietary polyphenols as modulators of brain functions: biological actions and molecular mechanisms underpinning their beneficial effects. Oxid Med Cell Longev. 2012;2012:914273. doi:10.1155/2012/914273
- 28. Khoshimov N.N; Raimova G.M.; Nasirov K.E.; Rakhimov R.N.; Azizov V.G. "The Effect of Sp-6 On The Transport of Mediators of NMDA-Receptors and Ca<sup>2+</sup>- channels in Synaptosomes of rat brain. "European Journal of Molecular & Clinical Medicine", 7, 3, 2020, 2435-2446.
- Campos-Esparza MR, Sánchez-Gómez MV, Matute C. Molecular mechanisms of neuroprotection by two natural antioxidant polyphenols. Cell Calcium.
   2009 Apr;45(4):358-68. doi: 10.1016/j.ceca.2008.12.007. Epub 2009 Feb 7. PMID: 19201465.
- 30. Khoshimov N.N., Rakhimov R.N., Akhmedova G.B., Azizov V.G. Investigation of the effect of polyphenol euphorbin on the transport of L Glutamate and calcium channels to synaptosomes of rat brain // European Journal of Medicine. – 2018. –

№. 6. – C. 72-82. DOI: 10.13187/ejm.2018.2.72

- 31. "European Convention for the Protection of Vertebrate Animals used for Experimental and other Scientific Purposes." Strasbourg. 1986. Available from: http://conventions.coe.int
- 32. Weiler, M. H., Gundersen C. B. and Jenden
  D. J. (1981) Choline uptake and acetylcholine synthesis in synaptosomes: Investigations using two differently labelled variants of choline//Neurochem.
  36. P.1802-1812.
- 33. Grynkiewicz G., Poenie M., Tsien R.Y. 'A new generation of Ca<sup>2+</sup>, indicators with greatly improved fluorescence properties'
  J. Biol. Chem. Vol. 260, 1985, P. 3440 3450.